# Mammalian valyl-tRNA synthetase forms a complex with the first elongation factor

Yu.A. Motorin, A.D. Wolfson, A.F. Orlovsky and K.L. Gladilin

A.N. Bakh Institute of Biochemistry, Academy of Sciences of the USSR, Moscow 117071, USSR

Received 2 August 1988

The high-molecular-mass form of valyl-tRNA synthetase is associated with the first elongation factor activity. It includes two polypeptides of about 50 kDa and two others of 40 and 30 kDa, identified as  $\alpha$ ,  $\beta$ ,  $\gamma$  and  $\delta$  subunits of eEF-1H. The complex of valyl-tRNA synthetase with eEF-1H is suggested to be a novel form of the first elongation factor.

valyl-tRNA synthetase; First elongation factor; High-molecular-mass complex

### 1. INTRODUCTION

One of the characteristic features of eukaryotic aminoacyl-tRNA synthetases is their ability to form high-molecular-mass complexes. The complex of nine aminoacyl-tRNA synthetases is now well characterised [1]. Valyl-tRNA synthetase is the only aminoacyl-tRNA synthetase, which exists in a high-molecular-mass form different from this complex [1-3]. Recently we succeeded in purifying the valvl-tRNA synthetase complex from rabbit liver [4]. This complex consists of polypeptides with molecular masses of 130, 50, 40 and 30 kDa and has a molecular mass of about 800 kDa. A complex with the same polypeptide composition was also isolated from rabbit liver by Waller et al. [5] and several other mammalian sources in our laboratory. The 130 kDa polypeptide was identified as valyl-tRNA synthetase. In this work we have identified the low-molecular-mass polypeptides of the complex as subunits of the heavy form of the first elongation factor (eEF-1H).

### 2. MATERIALS AND METHODS

Preparations of eEF-1 and eEF-2 were the kind gift of A.G.

Correspondence address: A.D. Wolfson, A.N. Bakh Institute of Biochemistry, Leninsky pr. 33, Moscow 117071, USSR

Ryasanov (Institute of Protein Research, Puschino). 80 S ribosomes from rabbit reticulocytes were isolated by a conventional procedure [6].

The valyl-tRNA synthetase complex was purified by gel filtration on Sepharose CL-6B and chromatography on Mono S and Mono Q columns (Pharmacia) as described previously [4]. The activity of valyl-tRNA synthetase was determined as described in [7].

EF-1 activity was assayed with the poly(U)-dependent translational system in 50  $\mu$ l of a mixture which contained 20 mM Tris-HCl (pH 7.6), 10 mM MgCl<sub>2</sub>, 100 mM KCl, 2 mM DTT, 1.2 mM GTP, 0.1 mg/ml poly(U), 10 pmol [<sup>14</sup>C]PhetRNA, 0.2  $A_{260}$  units of ribosomes and 1  $\mu$ g of EF-2.

Analytical gel filtration was performed on a Superose 6 HR 10/30 column (Pharmacia) in 25 mM potassium phosphate (pH 7.6), 1 mM MgCl<sub>2</sub>, 300 mM KCl, 2 mM  $\beta$ -mercaptoethanol, 10% glycerol. The flow rate was 0.5 ml/min and the sample volume 200  $\mu$ l.

SDS-gel electrophoresis and two-dimensional gel electrophoresis were performed on minigels with PhastSystem (Pharmacia) according to Pharmacia recommendations [8,9]. Gels were stained with silver according to [10].

#### 3. RESULTS AND DISCUSSION

Apart from the 130 kDa polypeptide identified earlier as valyl-tRNA synthetase, the complex contains polypeptides of 50, 40 and 30 kDa according to SDS electrophoresis. Since the SDS-electrophoresis pattern of the complex closely resembled that of eEF-1H (fig.1), we have suggested that the valyl-tRNA synthetase forms a

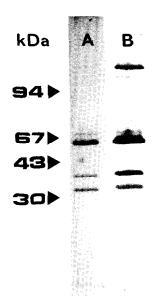


Fig.1. SDS gel electrophoresis on PhastGradient Gel 10-15. (A) eEF-1H from rabbit reticulocytes, (B) preparation of rabbit liver valyl-tRNA synthetase complex. Arrowheads indicate the position of molecular mass markers.

complex with eEF-1H. In order to prove this hypothesis several experiments were carried out.

The eEF-1 activity of the purified complex was assayed in the poly(U)-dependent translational system without eEF-1. The activity of the valyl-tRNA synthetase complex was approximately the

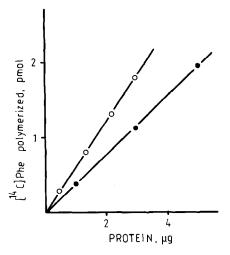


Fig.2. Stimulation of polyphenylalanine synthesis in poly(U)-dependent translational system. eEF-1H (0); valyl-tRNA synthetase complex (•).

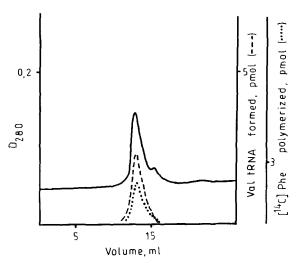


Fig.3. Gel filtration of the purified valyl-tRNA synthetase complex.

same as the conventional eEF-1H preparation (fig.2). The activities of eEF-1 and valyl-tRNA synthetase coelute upon gel filtration of the

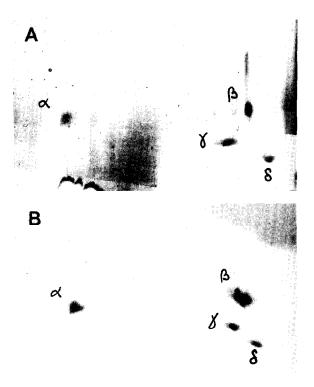


Fig. 4. Two-dimensional gel electrophoresis of valyl-tRNA synthetase complex (a) and of eEF-1H (b) from rabbit reticulocytes.

purified complex on Superose 6 (fig.3). The coelution of both activities was also observed during the isolation procedure (not shown). The valyl-tRNA synthetase complex was analysed by twodimensional gel electrophoresis for the identification of its polypeptides as subunits of eEF-1H (fig.4). Two polypeptides of about 50 kDa and pIvalues of about 9.0 and 5.0 are present in the complex (fig.4a). The comparison of the twodimensional electrophoresis data for the complex and eEF-1H shows that they are practically identical (fig.4). All the subunits of eEF-1H ( $\alpha$ ,  $\beta$ ,  $\gamma$ and  $\delta$  [11]) are present in the complex. The spot corresponding to valyl-tRNA synthetase was not observed on two-dimensional gels probably due to the high proteolytic lability of this enzyme.

Since all valyl-tRNA synthetase in the extracts of various eukaryotic cells exists exclusively in high-molecular-mass form ([2-4] and unpublished) and is tightly associated with the subunits of eEF-1H, the high-molecular-mass complex is probably a special form of EF-1, which exists in the extracts as well as 400 kDa eEF-1H and 50 kDa eEF-1L. Further studies will provide understanding of the function of this particular form of elongation factor.

Acknowledgement: The authors are grateful to A.G. Ryasanov for the helpful discussion.

## **REFERENCES**

- [1] Girakoglu, B., Mirande, M. and Waller, J.-P. (1985) FEBS Lett. 183, 185-190.
- [2] Mirande, M., Le Corre, D. and Waller, J.-P. (1985) Eur. J. Biochem. 147, 281-289.
- [3] Godar, D.E. and Yang, D.C.H. (1988) Biochemistry 27, 2181-2186.
- [4] Motorin, Yu.A., Wolfson, A.D., Orlovsky, A.F. and Gladilin, K.L. (1987) FEBS Lett. 220, 363-365.
- [5] Mirande, M., Kerjan, P., Lazard, M., Martinez, R., Bec, G. and Waller, J.-P. (1988) 14th International Congress of Biochemistry, Abstracts, S20-8.
- [6] Merrick, W.C. (1979) Methods Enzymol. 60, 108-129.
- [7] Wolfson, A.D., Motorin, Yu.A., Orlovsky, A.F. and Gladilin, K.L. (1987) Biokhimiya 52, 1847-1854.
- [8] PhastSystem Manual, Pharmacia LKB Publication.
- [9] Jagersten, C. and Edstrom, A. (1987) Proc. Scand-Elpho '87.
- [10] Gorg, A., Postel, W., Weser, J., Schivara, H.W. and Boesken, W.H. (1985) Science Tools 32, 5-9.
- [11] Carvalho, J.F., Carvalho, M.G.C. and Merrick, W.C. (1984) Arch. Biochem. Biophys. 234, 591-602.